

DescTools

A Hardworking Assistant for Describing Data

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Abstract

R sometimes makes ordinary tasks difficult. Virtually every data analysis project starts with describing data. The first thing to do will often be calculating summary statistics for all variables while listing the occurrence of nonresponse and missing data and producing some kind of graphics. This is a three-click process in SPSS, but regardless of the normality of this task, base R does not contain higher level functions for quickly describing huge datasets (meant regarding the number of variables, not records) adequately in a more or less automated way. There are facilities like `summary`, `describe` (`Hmisc`), `stat.desc` (`library pastecs`), but all of them are lacking some functionality or flexibility we would have expected.

Another point is, that there are quite a few commonly used functions, which curiously are not present in the *stats* package, think e.g. of skewness, kurtosis but also the Gini-coefficient, Cohen's Kappa or Somers' delta. This led to a rank growth of libraries implementing just one specific missing thing. There are plenty of "misc"-libraries out there, containing such functions and tests. We would normally end up using a dozen libraries, each time using just one single function out of it and suffering huge variety concerning NA-handling, recycling rules and so on.

R has been developed in a university environment. This will be clear at the latest then when you find yourself working in an office of an insurance and you realize that only MS-Office (and no LATEX) is installed on your system (and the IT guys won't give you admin rights). We were forced in this situation to write code for doing our reporting in MS-Word. (This works quite well for Windows, but not for Mac unfortunately.)

The first version of "DescTools" arose after completion of a project, where we had to describe a dataset under deadline pressure, and we started to gather our newly created functions and put them together.

This collection has meanwhile grown to a considerably versatile toolset for descriptive statistics, providing rich univariate and bivariate descriptions of data without expecting the user to say much.

There are numerous basic statistic functions and tests, possibly flexible and enriched with different approaches (if existing). Confidence intervals are extensively provided.

Recognizing that most problems can be satisfactorily visualized with bar-, scatter- and dotplots, still some more specific plot types are used in special cases and thus included in the library. Some of them are rather new, and some of them are based on types found scattered in the myriads of R-packages found out there (partly rewritten to meet the design goals of the package).

This document describes quickly the essentials of the package DescTools.



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Describing a data.frame

The function Desc is designed to describe variables depending on their type with some reasonable statistic measures and an adequate graphic representation. It includes code for describing

- logical variables
- factors, ordered and unordered
- integer variables (typically counts)
- numeric variables
- dates
- tables and matrices

Data frames will be split into their variables and the single variable will be described. A formula interface is implemented allowing to describe variables in dependence of others.

The output can either be sent to the R-console or as well directly redirected to a MS-Word document. The latter works only in Windows with MS-Office installed, but Mac users can leave the wrd argument away and add a plotit = TRUE argument to have the full results in the console.

Let's start with a quick description of some variables out of the integrated data.frame d.pizza.

```
library(DescTools)

# the results (and the plots) will either be displayed in the console
Desc(d.pizza[,c("driver", "temperature", "count", "weekday", "wine_ordered", "date")], plotit=TRUE)

# ... or we can start a new word instance and send the results directly to a word document
wrd <- GetNewWrd()
Desc(d.pizza[,c("driver", "temperature", "count", "weekday", "wine_ordered", "date")], wrd=wrd)
```

```
'data.frame':  1209 obs. of  4 variables:
 1 $ driver      : Factor w/ 7 levels "Butcher","Carpenter",...: 7 1 1 7 3 7 7 7 3 ...
 2 $ temperature : num  53 56.4 36.5 NA 50 27 33.9 54.8 48 54.4 ...
 3 $ count       : int   5 2 3 2 5 1 4 NA 3 6 ...
 4 $ weekday     : num   6 6 6 6 6 6 6 6 6 6 ...
 5 $ wine_ordered: int   0 0 0 0 0 0 1 NA 0 1 ...
 6 $ date        : Date, format: "2014-03-01" "2014-03-01" "2014-03-01" "2014-03-01" ...
```

First a simple Str() of the data.frame is performed. The result is no more than that of a str() command, extended with an enumeration of the variables.

After that, every single variable will be described according to the type of its class.

The first variable is an unordered factor. Factors are typically best described by a frequency table of their levels. The default order of the output table is following a pareto rule, the most frequent levels first.

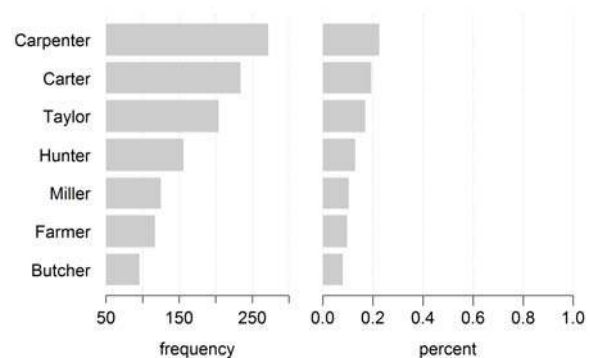
Ordered factors would be sorted after their natural order by default. The default order can be changed by setting the ord argument to either "desc", "asc", "name", "level".

The frequency table is truncated in the case that there are more than a dozen values (this can be avoided by setting the argument maxrows=NA, see: ?Desc.factor for more details).

1 : driver (factor)

length	n	NAs	levels	unique	dupes
1'209	1'204	5	7	7	y

	level	freq	perc	cumfreq	cumperc
1	Carpenter	272	.226	272	.226
2	Carter	234	.194	506	.420
3	Taylor	204	.169	710	.590
4	Hunter	156	.130	866	.719
5	Miller	125	.104	991	.823
6	Farmer	117	.097	1108	.920
7	Butcher	96	.080	1204	1.000



Synopsis

length	total number of elements in the vector, NAs are included here
n	number of valid cases, NAs, NaNs, Inf etc. are not counted here
NAs	number of missing values
levels	number of levels
unique	number of unique values.
	Note: This is not necessarily the same value as levels, as there might be empty levels. Thus the number of levels might be higher than the number of unique values (but not the other way round).
dupes	y(es) or n(o), reporting if there are any duplicate values in the vector. If "n" (for no) then there are only unique values in the variable.
freq	the count (absolute frequency) of the specific level. The order of a factors frequency table is by default chosen as "absolute frequency-decreasing".
perc	the relative frequency of the specific level
cumfreq	the cumulative frequencies of the levels
cumperc	the same for the percentage values

The next variable, the temperature of the delivered pizza, is numeric. Numeric variables are described by the most usual statistical measures for location, variation and shape.

2 : temperature (numeric)

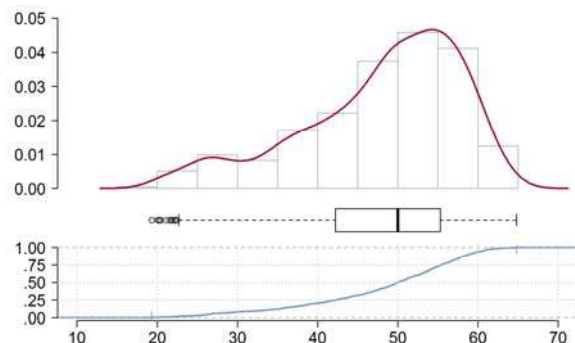
length	n	NAs	unique	0s	mean	meanSE
1'209	1'170	39	375	0	47.937	0.291

.05	.10	.25	median	.75	.90	.95
26.700	33.290	42.225	50	55.300	58.800	60.500

rng	sd	vcoef	mad	IQR	skew	kurt
45.500	9.938	0.207	9.192	13.075	-0.842	0.051

lowest : 19.3, 19.4, 20, 20.2 (2), 20.35
highest: 63.8, 64.1, 64.6, 64.7, 64.8

Shapiro-Wilks normality test p.value : < 2.22e-16



The first measures length, n, NAs, unique have the same meaning as above.

0s	total number of 0s, say zero values.
mean	the mean of the vector, NAs are silently removed.
meanSE	standard error of the mean, $sd(x) / \sqrt{n}$. this can be used to construct the confidence intervals for the mean, defined as $qt(p = 0.025, df = n-1) * sd(x) / \sqrt{n}$. (See also: function <code>MeanCI(...)</code>)
.05, ..., .95	quantiles of x, starting with 5%, 10%, 1. quartile, median etc.
rng	range of x, $\max(x) - \min(x)$
sd	standard deviation
vcoef	variation coefficient, defined as $sd(x) / \text{mean}(x)$
mad	median absolute deviation
IQR	inter quartiles range
skew	skewness of x
kurt	kurtosis of x
lowest	the smallest 5 values. Note that, if there are bindings here, the frequency of each value will be reported in brackets.
highest	same as lowest, but on the other end
Shapiro-Wilks	performs a Shapiro-Wilks normality test and reports its p-value. Shapiro-Wilks will be replaced by the Anderson-Darling-Test, if $\text{length}(x) > 5000$.
plot	the plot combines a histogram with a density plot, a boxplot and the ecdf-plot, as produced by the function <code>PlotFdist</code> .

The next variable is a count variable, whose nature is somewhat between numeric and factors as far as descriptive measures are concerned. In fact, if there are only just a few unique values, then the factor representation (frequencies) might be more appropriate than the numeric description (with densities etc.). We draw the line between factor and numeric representation at a dozen of unique values in x. Beyond that number, the numeric description will be reported and for fewer values the factor representation will be used.

3 : count (integer)

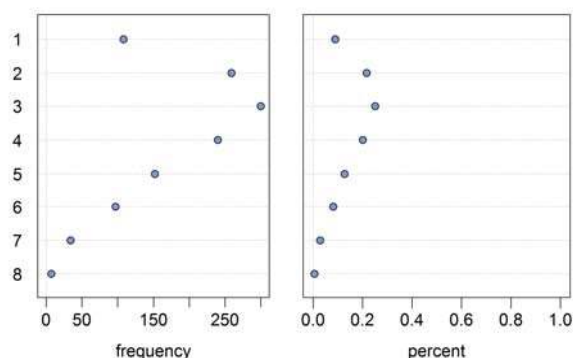
```
length      n    NAs unique    0s    mean meanSE
1'209 1'197    12     8      0  3.444  0.045

.05    .10    .25 median    .75    .90    .95
1      2      2      3      4      6      6

rng      sd    vcoef    mad    IQR    skew    kurt
7  1.556  0.452  1.483    2  0.454 -0.363

Shapiro-Wilks normality test  p.value : < 2.22e-16
```

	level	freq	perc	cumfreq	cumperc
1	1	108	.090	108	.090
2	2	259	.216	367	.307
3	3	300	.251	667	.557
4	4	240	.201	907	.758
5	5	152	.127	1059	.885
6	6	97	.081	1156	.966
7	7	34	.028	1190	.994
8	8	7	.006	1197	1.000



plot the plot is produced as a (horizontal) dotchart. More than 12 unique values are truncated (a warning is placed in the plot area). Use maxrows argument to override this default (NA for all).

Two dotcharts are created, the left one shows the absolute frequencies, the right one the percentages. On the left plot the x-axis might be adapted to the data (as R does by default). The percentages will always be displayed on a 0:1-range.

The plot width is adapted to the length of the labels. If the labels get too long, they will be truncated and displayed with ellipsis (...).

If there's a numeric variable with only one or two handfuls of unique values then a description by means of a histogram and a density curve is not really adequate. Therefore we change the graphic representation from a histogram to a histogram like h-type plot without density curve.

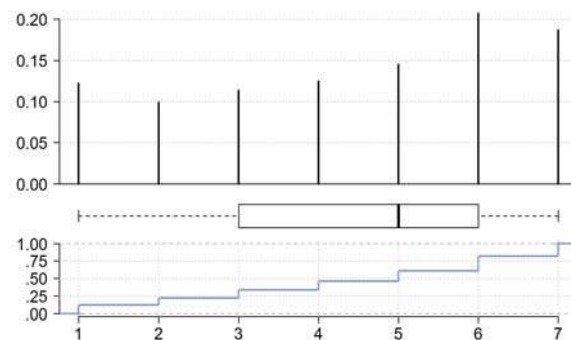
4 : weekday (numeric)

length	n	NAs	unique	0s	mean	meanSE
1'209	1'177	32	7	0	4.441	0.059

.05	.10	.25	median	.75	.90	.95
1	1	3	5	6	7	7

rng	sd	vcoef	mad	IQR	skew	kurt
6	2.019	0.455	2.965	3	-0.345	-1.170

lowest : 1 (144), 2 (117), 3 (134), 4 (147), 5 (171)
highest: 3 (134), 4 (147), 5 (171), 6 (244), 7 (220)



Shapiro-Wilks normality test p.value : < 2.22e-16

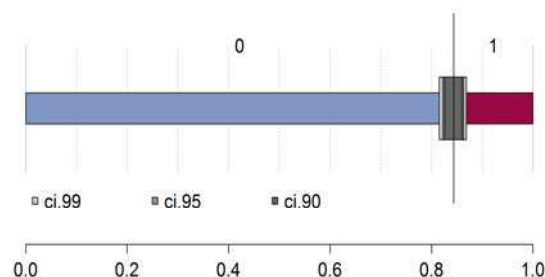
The variable wine_ordered contains only two values, 0 and 1. Dichotomous variables do not have real dense (univariate) information. But still it is interesting to know, how many NAs there are, besides the frequencies of course. The individual frequencies are reported together with a confidence interval, calculated by BinomCI using the option "Wilson".

5 : wine_ordered (integer)

length	n	NAs	unique
1'209	1'197	12	2

freq	perc	lci.95	uci.95 ¹
0 1010	.844	.822	.863
1 187	.156	.137	.178

¹ 95%-CI Wilson



plot this is basically a univariate horizontal stacked barplot, with confidence intervals on the confidence levels of 0.90, 0.95 and 0.99. The vertical line denominates the point estimator.

A date variable is harder to describe. What characteristics would one want to know from a date?
 We would normally choose a description similar to numeric values, supplemented by an analysis of the weekday and month.

6 : date (Date)

```
length      n    NAs unique
1'209  1'177     32     31
```

lowest : 2014-03-01 (42), 2014-03-02 (46), 2014-03-03 (26), 2014-03-04 (19)
 highest: 2014-03-28 (46), 2014-03-29 (53), 2014-03-30 (43), 2014-03-31 (34)

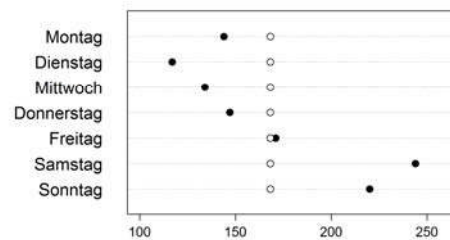
Weekdays:

	level	freq	perc	cumfreq	cumperc	exp	res
1	Montag	144	.122	144	.122	168.1	-1.9
2	Dienstag	117	.099	261	.222	168.1	-3.9
3	Mittwoch	134	.114	395	.336	168.1	-2.6
4	Donnerstag	147	.125	542	.460	168.1	-1.6
5	Freitag	171	.145	713	.606	168.1	.2
6	Samstag	244	.207	957	.813	168.1	5.9
7	Sonntag	220	.187	1177	1.000	168.1	4.0

Chi-squared test for given probabilities

data: table(xd)

X-squared = 78.8785, df = 6, p-value = 6.09e-15



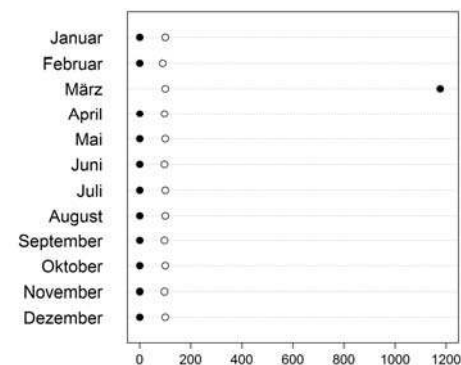
Months:

	level	freq	perc	cumfreq	cumperc	exp	prs.res
1	Januar	0	0	0	0	99.7	-10.0
2	Februar	0	0	0	0	93.3	-9.7
3	März	1177	1	1177	1	99.7	107.9
4	April	0	0	1177	1	96.5	-9.8
5	Mai	0	0	1177	1	99.7	-10.0
6	Juni	0	0	1177	1	96.5	-9.8
7	Juli	0	0	1177	1	99.7	-10.0
8	August	0	0	1177	1	99.7	-10.0
9	September	0	0	1177	1	96.5	-9.8
10	Oktober	0	0	1177	1	99.7	-10.0
11	November	0	0	1177	1	96.5	-9.8
12	Dezember	0	0	1177	1	99.7	-10.0

Chi-squared test for given probabilities

data: tab

X-squared = 12719.19, df = 11, p-value < 2.2e-16

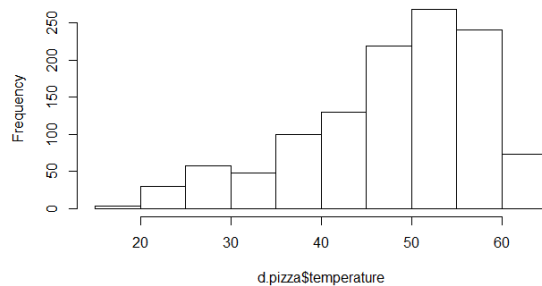


Simple Frequencies

There are as well more atomic functions in the package. The function `Freq` is designed to give the numeric representation of a histogram, produced by `hist`. It displays the frequencies and the percentages of a binned variable with the same logic as in `hist`. The single and cumulative frequencies values are reported.

```
Freq(d.pizza$temperature)
hist(d.pizza$temperature)
```

	level	freq	perc	cumfreq	cumperc
1	[15,20]	3	0.003	3	0.003
2	(20,25]	30	0.026	33	0.028
3	(25,30]	58	0.050	91	0.078
4	(30,35]	48	0.041	139	0.119
5	(35,40]	100	0.085	239	0.204
6	(40,45]	130	0.111	369	0.315
7	(45,50]	219	0.187	588	0.503
8	(50,55]	268	0.229	856	0.732
9	(55,60]	241	0.206	1097	0.938
10	(60,65]	73	0.062	1170	1.000



Pairwise descriptions

`Desc` implements a formula interface allowing to define bivariate descriptions straight forward.

A numeric variable vs. a categorical is best described by group wise measures. Here the valid pairs are reported first. Missing values in the single groups are documented in the results table and missing values on the grouping factor are mentioned with a warning at the end of the table, if existing at all.

```
Desc(temperature ~ driver, d.pizza, digits=1, plotit=TRUE)
```

temperature ~ driver(numeric ~ categorical)

Summary:

n pairs: 1'209, valid: 1'166 (96%), missings: 43 (4%), groups: 7

	Butcher	Carpenter	Carter	Farmer	Hunter	Miller	Taylor
mean	49.6	43.5 ¹	50.4	50.9	52.1 ²	47.5	45.1
median	51.4	44.8 ¹	51.8	54.1	55.1 ²	49.6	48.5
sd	8.8	9.4	8.5	9.0	8.9	8.9	11.4
IQR	12.0	12.5	11.3	11.2	11.6	8.8	18.4
n	96	253	226	117	156	121	197
np	0.082	0.217	0.194	0.100	0.134	0.104	0.169
NAs	0	19	8	0	0	4	7
0s	0	0	0	0	0	0	0

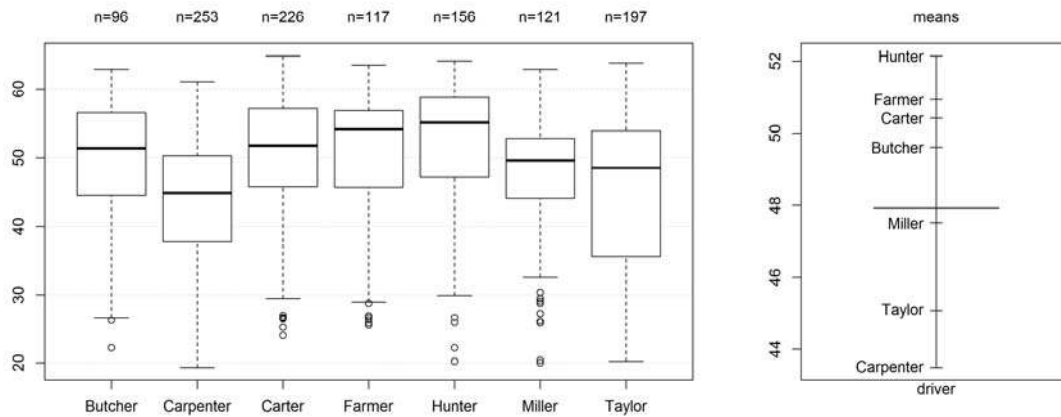
¹ min, ² max

Kruskal-Wallis rank sum test:

Kruskal-Wallis chi-squared = 141.9349, df = 6, p-value < 2.2e-16

Warning:

Grouping variable contains 5 NAs (0.414%).



plot a boxplot combined with a means-plot as used in anova.

Two categorical variables are described by a contingency table. Again the total pairs, the valid pairs and the missings are again reported first.

Two numerical variables have no obvious standard description as their relationship can have many forms. We report therefore only the simple correlation coefficients (Pearson, Spearman and Kendall).

The variables are plotted as xy-scatterplots with interchanging mutual dependency, supplemented with a LOESS smoother.

```
Desc(temperature ~ delivery_min, d.pizza, plotit=TRUE)
```

temperature ~ delivery_min (numeric ~ numeric)

Summary:

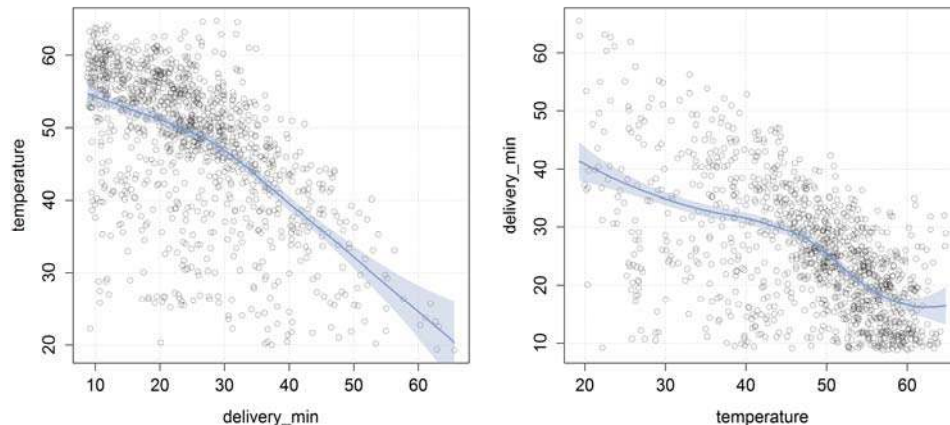
n pairs: 1'209, valid: 1'170 (97%), missings: 39 (3%)

Pearson corr. : -0.575

Spearman corr.: -0.573

Kendall corr. : -0.422

Scatterplots for two numeric variables:



Two categorical variables are described by a contingency table, as shown in the next chapter.

Tables

Get the frequencies and the percentages of a RxC-dimensional contingency table and output a flat table. Expected values and standardized residuals can be computed.

```
# A)
PercTable(d.pizza$driver, d.pizza$city, margins=c(1,2), rfrq="101")
# B)
PercTable(d.pizza$driver, d.pizza$city, margins=c(1,2), rfrq="000", expected=TRUE, stdres=TRUE,
          digits=1)
```

A) Frequencies and percentages

		Brent	Camden	Westminster	Sum
Butcher	freq	72	1	22	95
	perc	.060	.001	.018	.080
	p.col	.152	.003	.058	.080
Carpenter	freq	29	19	221	269
	perc	.024	.016	.185	.225
	p.col	.061	.056	.582	.225
Carter	freq	177	47	5	229
	perc	.148	.039	.004	.192
	p.col	.374	.138	.013	.192
Farmer	freq	19	87	11	117
	perc	.016	.073	.009	.098
	p.col	.040	.255	.029	.098
Hunter	freq	128	4	24	156
	perc	.107	.003	.020	.131
	p.col	.271	.012	.063	.131
Miller	freq	6	41	77	124
	perc	.005	.034	.064	.104
	p.col	.013	.120	.203	.104
Taylor	freq	42	142	20	204
	perc	.035	.119	.017	.171
	p.col	.089	.416	.053	.171
Sum	freq	473	341	380	1194
	perc	.396	.286	.318	1.000
	p.col	1.000	1.000	1.000	1.000

B) Expected values and std. residuals

		Brent	Camden	Westminster	Sum
Butcher	freq	72	1	22	95
	exp	37.6	27.1	30.2	.
	stdres	7.5	-6.2	-1.9	.
Carpenter	freq	29	19	221	269
	exp	106.6	76.8	85.6	.
	stdres	-11.0	-8.9	20.1	.
Carter	freq	177	47	5	229
	exp	90.7	65.4	72.9	.
	stdres	13.0	-3.0	-10.7	.
Farmer	freq	19	87	11	117
	exp	46.3	33.4	37.2	.
	stdres	-5.4	11.5	-5.5	.
Hunter	freq	128	4	24	156
	exp	61.8	44.6	49.6	.
	stdres	11.6	-7.7	-4.7	.
Miller	freq	6	41	77	124
	exp	49.1	35.4	39.5	.
	stdres	-8.4	1.2	7.6	.
Taylor	freq	42	142	20	204
	exp	80.8	58.3	64.9	.
	stdres	-6.1	14.3	-7.4	.
Sum	freq	473	341	380	1194
	exp
	stdres

There are quite a few suggestions for the description of tables out there. We use a mix between SAS- and SPSS-flavour here. Let's take a SAS-example¹ and describe the table with SPSS-verbosity:

```
pain <- as.table(matrix(c(26,26,23,18, 9,
                          6, 7, 9,14,23), nrow=2, byrow=TRUE,
                          dimnames=list(treat=c("No","Yes"), strength=0:4)) )
Desc(pain, verb="high", wrd=wrds)
```

Clinical Trial for Treatment of Pain (2x5-table)

Summary:

n: 161, rows: 2, columns: 5

Pearson's Chi-squared test:

X-squared = 26.6025, df = 4, p-value = 2.392e-05

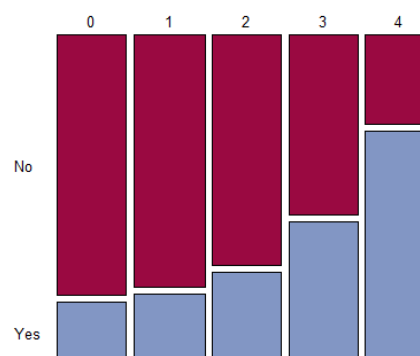
Likelihood Ratio:

X-squared = 26.6689, df = 4, p-value = 2.319e-05

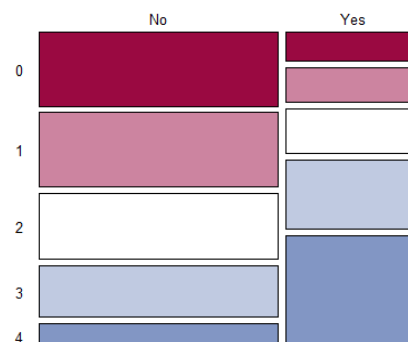
Mantel-Haenszel Chi-squared:

X-squared = 22.8188, df = 1, p-value = 1.78e-06

	estimate	lwr.ci	upr.ci
Phi Coeff.	.4065	-	-
Contingency Coeff.	.3766	-	-
Cramer V	.4065	.2716	.5636
Goodman Kruskal Gamma	.5313	.3480	.7146
Kendall Tau-b	.3373	.2114	.4631
Stuart Tau-c	.4111	.2547	.5675
Somers D C R	.4427	.2786	.6068
Somers D R C	.2569	.1593	.3546
Pearson Correlation	.3776	.2368	.5029
Spearman Correlation	.3771	.2362	.5024
Lambda C R	.1250	.0000	.2547
Lambda R C	.2373	.0732	.4014
Lambda sym	.1604	.0388	.2821
Uncertainty Coeff. C R	.0515	.0140	.0890
Uncertainty Coeff. R C	.1261	.0346	.2175
Uncertainty Coeff. sym	.0731	.0199	.1262
Mutual Information	.1195	-	-



		strength	0	1	2	3	4	Sum
No	freq		26	26	23	18	9	102
	perc		.161	.161	.143	.112	.056	.634
	p.row		.255	.255	.225	.176	.088	1.000
	p.col		.812	.788	.719	.562	.281	.634
Yes	freq		6	7	9	14	23	59
	perc		.037	.043	.056	.087	.143	.366
	p.row		.102	.119	.153	.237	.390	1.000
	p.col		.188	.212	.281	.438	.719	.366
Sum	freq		32	33	32	32	32	161
	perc		.199	.205	.199	.199	.199	1.000
	p.row		.199	.205	.199	.199	.199	1.000
	p.col		1.000	1.000	1.000	1.000	1.000	1.000



¹ <http://support.sas.com/documentation/cdl/en/statugfreq/63124/PDF/default/statugfreq.pdf>, S. 1821

Let's have a look at a 2x2-table. We extend the ChiSquare-test by the exact Fisher test and report McNemar's symmetry test. The odds ratio and relative risk are displayed in addition by default. The verbosity can be set to "medium", "low" and "high".

```
heart <- as.table(matrix(c(11,4,
                          2,6), nrow=2, byrow=TRUE,
                          dimnames=list(Cholesterol=c("High","Low"), Response=c("Yes","No"))))

Desc(heart, wrd=wrds, horiz=FALSE)
```

Heart (2x2-table)

Summary:

n: 23, rows: 2, columns: 2

Pearson's Chi-squared test:

X-squared = 3.1879, df = 1, p-value = 0.07418

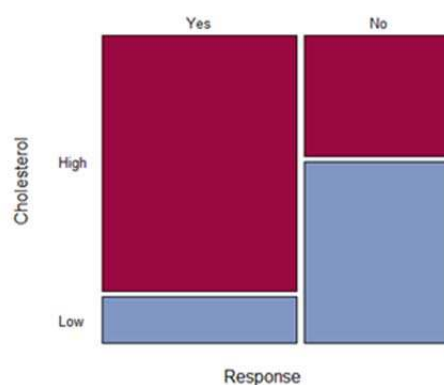
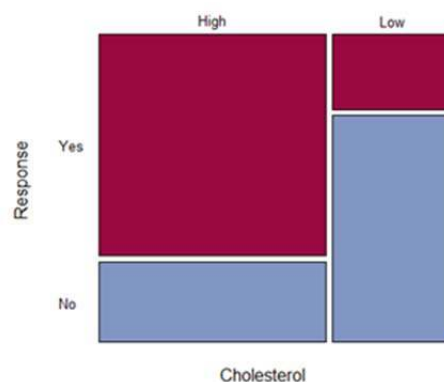
Fisher's exact test p-value = 0.03931

McNemar's chi-squared = 0.1667, df = 1, p-value = 0.6831

	estimate	lwr.ci	upr.ci
odds ratio	8.250	1.154	59.003
rel. risk (col1)	2.933	0.850	10.120
rel. risk (col2)	0.356	0.140	0.901

Phi-Coefficient	0.464
Contingency Coeff.	0.421
Cramer's V	0.464

		Response	Yes	No	Sum
Cholesterol					
High	freq		11	4	15
	perc		.478	.174	.652
	p.row		.733	.267	1.000
	p.col		.846	.400	.652
Low	freq		2	6	8
	perc		.087	.261	.348
	p.row		.250	.750	1.000
	p.col		.154	.600	.348
Sum	freq		13	10	23
	perc		.565	.435	1.000
	p.row		.565	.435	1.000
	p.col		1.000	1.000	1.000



Warning message:

In chisq.test(x) : Chi-squared approximation may be incorrect

Lorenz curves

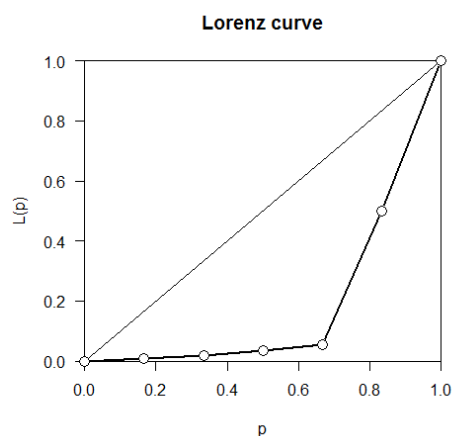
Lorenz-curves can be found in other libraries. This implementation starts with that from the library `ineq`, adding some value by calculating confidence intervals for the Gini coefficient.

```
x <- c(10, 10, 20, 20, 500, 560)

lc <- Lc(x)
plot(lc)
points(lc$p, lc$L, cex=1.5, pch=21, bg="white", col="black", xpd=TRUE)

Gini(x)
Gini(x, unbiased = FALSE)

Gini(x, conf.level = 0.95)
```



```
> Gini(x)
[1] 0.7535714

> Gini(x, unbiased = FALSE)
[1] 0.6279762

> Gini(x, conf.level=0.95)
      gini   lwr.ci   upr.ci
0.7535714 0.2000000 0.8967742
```

Lineplots

There are many flavours of line plots. Most (all?) of them can be handled by the function `matplot`. We generally desist from defining own functions, that only set suitable arguments for another already existing function, as we fear we would run into a forest of new functions, loosing overview.

Yet the parametrization of `matplot` can be a haunting experience and so we integrate some common examples here in the sense of a “How-To” tutorial.

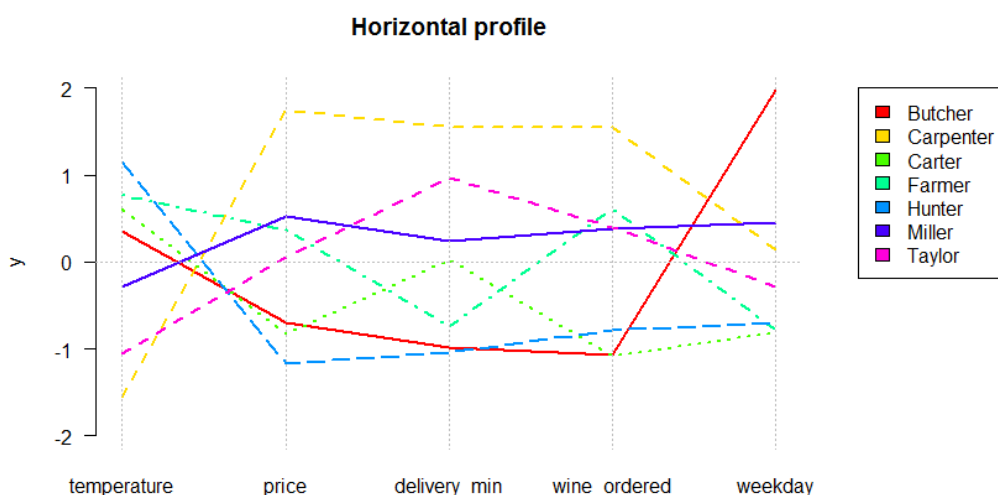
Let’s for example have a horizontal profile of the driver’s characteristics.

```
m <- data.frame(lapply(d.pizza[,c("temperature","price","delivery_min","wine_ordered","weekday")],
  tapply, d.pizza$driver, mean, na.rm=TRUE))
(ms <- data.frame(lapply(m, scale)))      # lets scale that
```

	temperature	price	delivery_min	wine_ordered	weekday
Butcher	0.3605689	-0.69917381	-0.98046684	-1.0738446	1.9826284
Carpenter	-1.5481318	1.74805901	1.54851320	1.5445402	0.1389367
Carter	0.6105633	-0.82596309	0.02841316	-1.0840337	-0.8062020
Farmer	0.7718643	0.36562860	-0.74842415	0.6105001	-0.7800183
Hunter	1.1473246	-1.16829499	-1.04738479	-0.7792855	-0.7038441
Miller	-0.2918676	0.52072004	0.23662429	0.3794541	0.4596817
Taylor	-1.0503216	0.05902424	0.96272512	0.4026695	-0.2911825

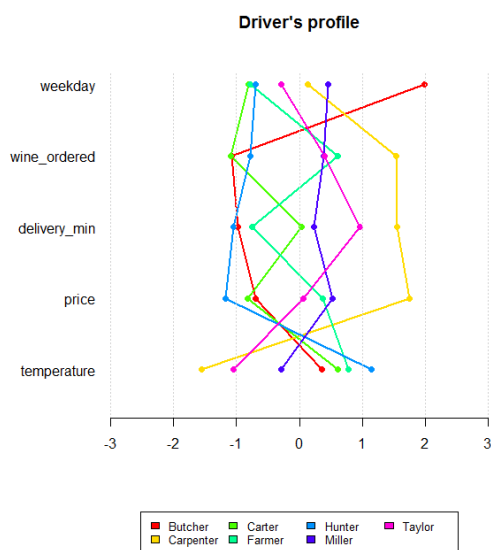
```
x <- 1:ncol(ms)
y <- t(ms)

windows(8.8,5)
par(mar=c(5,4,4,10)+.1)
matplot(x, y, type="l", col=rainbow(nrow(ms)), xaxt="n", las=1, lwd=2, frame.plot=FALSE, ylim=c(-2,2),
  xlab="", main="Horizontal profile")
abline(h=0, v=1:5, lty="dotted", col="grey")
par(xpd=TRUE)
legend(x=5.5, y=2, legend=rownames(ms), fill=rainbow(nrow(ms)))
axis(side=1, at=1:5, labels=colnames(ms), las=1, col="white")
```

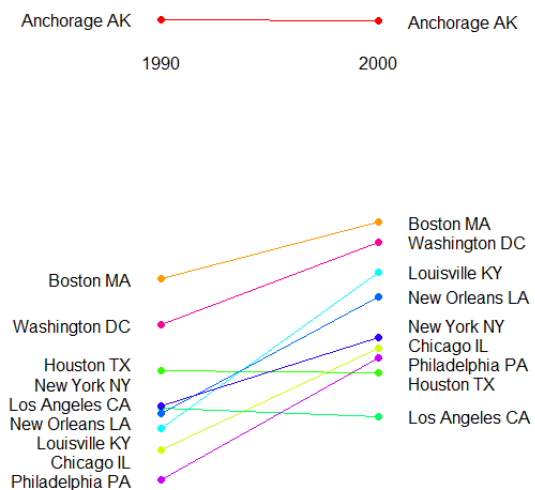


And the same, but on the vertical axis. (A)

```
par(mar=c(8,8,5,2))
matplot(x=y, y=x, type="l", pch=1:5, frame.plot=FALSE, axes=FALSE, xlab="", ylab="", lty="solid",
        col=rainbow(nrow(ms)), xlim=c(-3,3), ylim=c(0.5,ncol(ms)), main="Driver's profile", lwd=2)
matpoints(x=y, y=x, col=rainbow(nrow(ms)), pch=16)
grid(ny=NA)
axis(side=1, las=1)
mtext(colnames(ms), side=2, at=1:ncol(ms), las=2)
par(xpd=TRUE)
legend(x=0, y=-1, legend=rownames(ms), fill=rainbow(nrow(ms)), xjust=0.5, ncol=4, cex=0.8)
```



A)



B)

“Bumpchart”

Plot B is sometimes called bumpchart (Jim Lemon).

```
# example from plotrix (bumpchart)
edu <- matrix(c(90.4,90.3,75.7,78.9,66,71.8,70.5,70.4,68.4,67.9,
               67.2,76.1,68.1,74.7,68.5,72.4,64.3,71.2,73.1,77.8), ncol=2, byrow=TRUE)
rownames(edu) <- c("Anchorage AK","Boston MA","Chicago IL",
                  "Houston TX","Los Angeles CA","Louisville KY","New Orleans LA",
                  "New York NY","Philadelphia PA","Washington DC")
colnames(edu) <- c(1990,2000)

par(mar=c(5,10,5,10))
matplot(x=1:2, y=t(edu), type="l", frame.plot=FALSE, axes=FALSE, xlab="",
        ylab="", lty="solid", col=rainbow(10))
matpoints(x=1:2, y=t(edu), pch=16, frame.plot=FALSE, axes=FALSE, xlab="",
          ylab="", lty="solid", col=rainbow(10))

sapply( 1:2, function(i) mtext(rownames(edu), side=2*i,
                               at=SpreadOut(edu[,i], mindist=1.1), line=1, las=1 ))
mtext(colnames(edu), side=3, at=1:2, line=-3.5, las=1 )
```

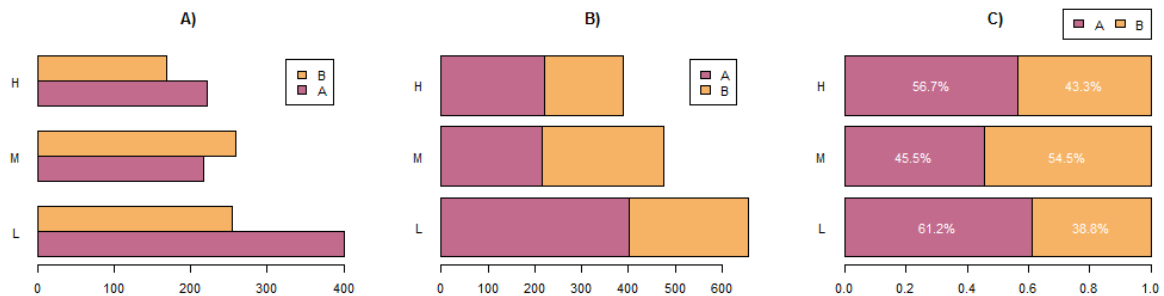
Barplot horizontal

A simple barplot, once with absolute values, once with percentages.

```
windows(height=3, width=11); par(mfrow=c(1,3))
col <- SetAlpha(PalHelsana(), 0.6)
tab <- matrix(c(401,216,221,254,259,169), nrow=2, byrow=TRUE,
  dimnames=list(wool=c("A","B"), tension=c("L","M","H")))
ptab <- prop.table(tab, 2)

# A)
barplot(tab, beside = TRUE, horiz=TRUE, main="A"),
  col = col[1:2], las = 1, legend = rownames(tab))
# B)
barplot(tab, beside = FALSE, horiz=TRUE, main="B"),
  col = col[1:2], las = 1, legend = rownames(tab))
# C)
b <- barplot(ptab, beside = FALSE, horiz=TRUE, main="C"),
  col = col[1:2], las = 1, legend.text = rownames(tab),
  args.legend = list(x=1, y=4.4, bg="white", ncol=2))

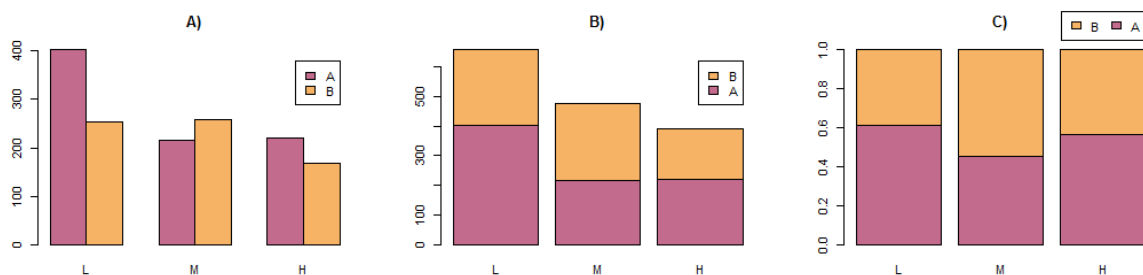
x <- t(apply(ptab, 2, Midx, incl.zero=TRUE, cumulate=TRUE))
text(paste(round(t(ptab),3) * 100, "%",sep=""), x=x, y=b, col="white")
```



Barplot vertical

This same as above but with vertical bars.

```
# A)
barplot(tab, beside = TRUE, main="A"),
  col = col[1:2], legend = rownames(tab))
# B)
barplot(tab, beside = FALSE, main="B"),
  col = col, legend = rownames(tab))
# C)
barplot(ptab, beside = FALSE, main="C"),
  col = col, legend.text = rownames(tab),
  args.legend = list(x=3.6, y=1.2, bg="white", ncol=2))
```



Barplot (specials)

Some specials like overlapping bars, connecting lines or error bars in combination with a barplot.

```

windows(height=3,11)
par(mfrow=c(1,3))

# A) Overlapping bars -----
blue <- rbind(c(5, 3, 4, 3),
              c(3, 2, 5, 1))
dimnames(blue) <- list(c("A","B"),c("t1","t2","t3","t4"))
red <- rbind(c(1.7,3.5,1.6,1.1),
             c(2.1,1.0,1.7,0.5))
dimnames(red) <- list(c("A","B"),c("t1","t2","t3","t4"))

# Set parameters
osp <- 0.5          # overlapping part in %
sp <- 1            # spacing between the bars

nbars <- dim(blue)[2] # how many bars do we have?

# Create first barplot
b <- barplot( blue, col=c("lightblue","blue"), main="A")
             , beside=FALSE, ylim=c(0,10), axisnames=FALSE
             , xlim=c(0, nbars*2-osp)      # enlarge x-Axis
             , space=c(0, rep(sp, nbars-1)) # set spacing=1, starting with 0
             )

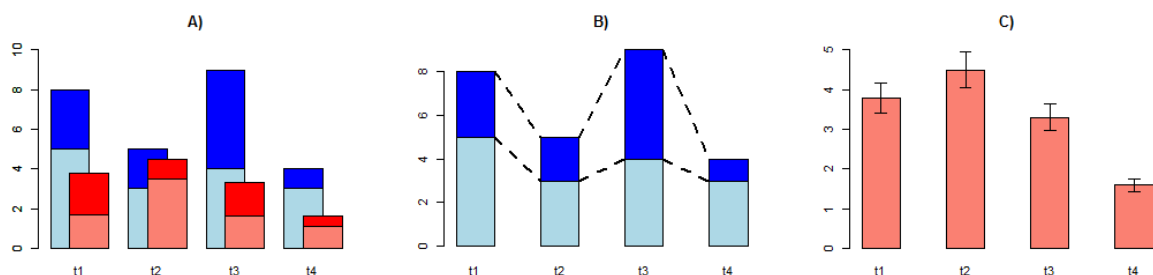
# Draw the red series
barplot( red, col=c("salmon","red"), beside=FALSE
        , space=c(1-osp, rep(1, nbars-1)) # shift to right by 1-osp
        , axisnames=FALSE, add=TRUE)

# Create axis separately, such that labels can be shifted to the left
axis(1, labels=colnames(red), at=b+(1-osp)/2, tick=FALSE, las=1)

# B) Connecting lines -----
barplot(blue, col=c("lightblue","blue"), space=1.2, main="B") )
ConnLines(blue, lwd=2, lty="dashed", space=1.2)

# C) Add error bars -----
cred <- apply(red, 2, sum)
b <- barplot(cred, col=c("salmon"), space=1.2, ylim=c(0,5), main="C") )
ErrBars(from=cred * .90, to=cred * 1.1, pos=b)

```



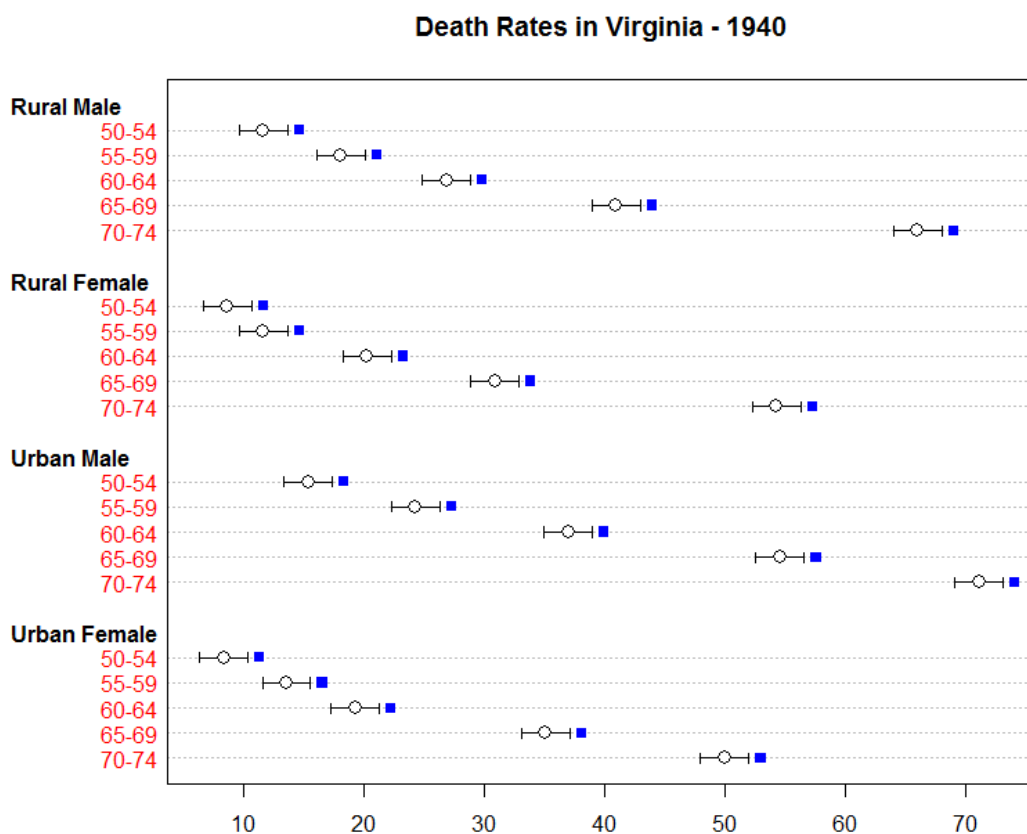
PlotDot

The base function `dotchart` has been improved but still has some potential for extensions. Especially an `add` argument is sometimes useful and returning the y-coordinates for the points would allow to add elements.

`PlotDot` implements these extensions and allows adding error bars. This is interesting, as the calculation of the x-limits should be done with respect to the bars and not only to the points.

```
# add some error bars
PlotDot(VADeaths, main="Death Rates in Virginia - 1940", col="red", pch=NA,
        args.errbars = list(from=VADeaths-2, to=VADeaths+2, mid=VADeaths,
                             pch=21, cex=1.4))

# add some other values
PlotDot(VADeaths+3, pch=15, col="blue", add=TRUE, labels=NA)
```



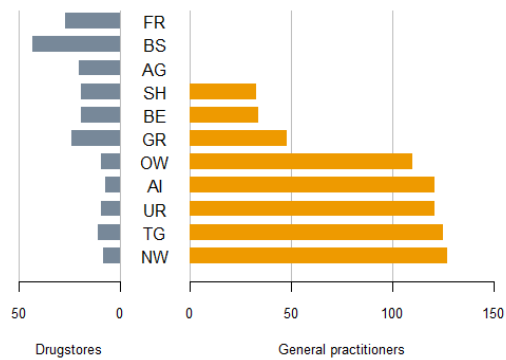
PlotPyramid

A special kind of horizontal barplot is a “pyramid plot”, where the bars are plotted back to back. This is sometimes needed, when your boss has specific and strict ideas how his presentation should look like.

```
d.sda <- data.frame(
  kt_x = c("NW","TG","UR","AI","OW","GR","BE","SH","AG","BS","FR"),
  apo_n = c( 8, 11,  9,  7,  9, 24, 19, 19, 20, 43, 27 ),
  sda_n = c(127, 125, 121, 121, 110, 48, 34, 33,  0,  0,  0 ))

PlotPyramid(lx=d.sda[,c("apo_n","sda_n")], ylab=d.sda$kt_x,
  col=c("lightslategray", "orange2"), border = NA, ylab.x=0, xlim=c(-110,250),
  gapwidth = NULL, cex.lab = 0.8, cex.axis=0.8, xaxt = TRUE,
  lxlab="Drugstores", rxlab="General practitioners",
  main="Density of general practitioners and drugstores",
  space=0.5, args.grid=list(lty=1))
```

Density of general practitioners and drugstores



Areaplot

Areaplots have a high “ink factor”², say they use much ink to display the information and are therefore rarely the best way of representing data. But again, when your boss wants it this way, here’s a function to produce it easily.

```
t.oil <- t(matrix(c(13.3,11.4, 9.7,10.6,12.7,11.0,10.6,13.5,
                  5.3, 3.6, 5.8, 8.4, 9.1,14.8,10.6, 9.6,
                  4.9, 3.1, 3.0, 6.0,12.2, 7.1, 7.3,10.0,
                  2.1, 2.6, 2.7, 3.5, 4.7, 5.0, 4.4, 4.3), nrow=4, byrow=TRUE,
              dimnames = list(c("ExxonMobil", "BP", "Shell", "Eni"),
                              c("1998", "1999", "2000", "2001", "2002", "2003", "2004", "2005"))))
t(t.oil)

par(mfrow=c(1,2), mar=c(5,4,5,5))
col <- SetAlpha(PalHelsana(), 0.7)
PlotArea(t.oil, col = col, las = 1, frame.plot=FALSE)
mtext(side=4, text=colnames(t.oil), las=1,
      at=Midx(tail(t.oil, 1)[,], incl.zero=TRUE, cumulate=TRUE))

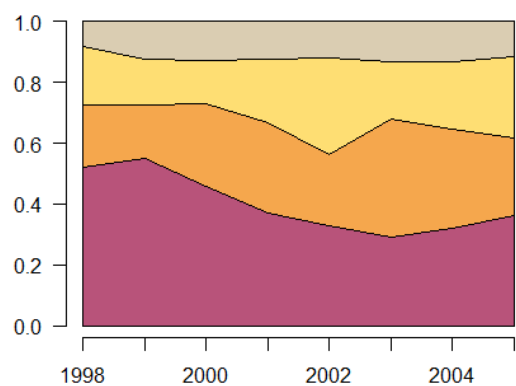
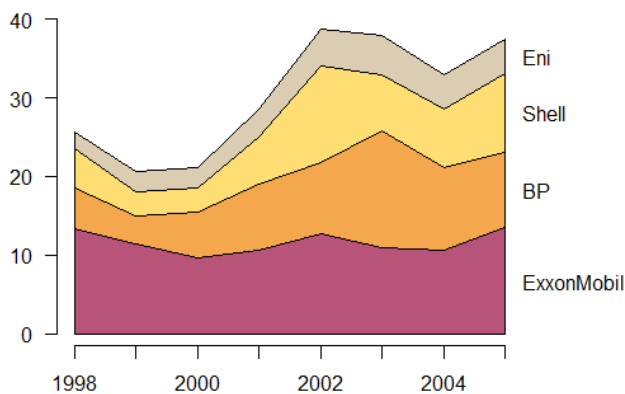
PlotArea(prop.table(t.oil, 1), col = col, las = 1, frame.plot=FALSE)
```

tab (absolute values)

```
> t(t.oil)
      1998 1999 2000 2001 2002 2003 2004 2005
ExxonMobil 13.3 11.4  9.7 10.6 12.7 11.0 10.6 13.5
BP          5.3  3.6  5.8  8.4  9.1 14.8 10.6  9.6
Shell       4.9  3.1  3.0  6.0 12.2  7.1  7.3 10.0
Eni         2.1  2.6  2.7  3.5  4.7  5.0  4.4  4.3
```

ptab (relative values)

```
      1998 1999 2000 2001 2002 2003 2004 2005
ExxonMobil 0.520 0.551 0.458 0.372 0.328 0.290 0.322 0.361
BP          0.207 0.174 0.274 0.295 0.235 0.391 0.322 0.257
Shell       0.191 0.150 0.142 0.211 0.315 0.187 0.222 0.267
Eni         0.082 0.126 0.127 0.123 0.121 0.132 0.134 0.115
```



² Tufte, Edward R (2001) [1983], The Visual Display of Quantitative Information (2nd ed.), Cheshire, CT: Graphics Press, ISBN 0-9613921-4-2.

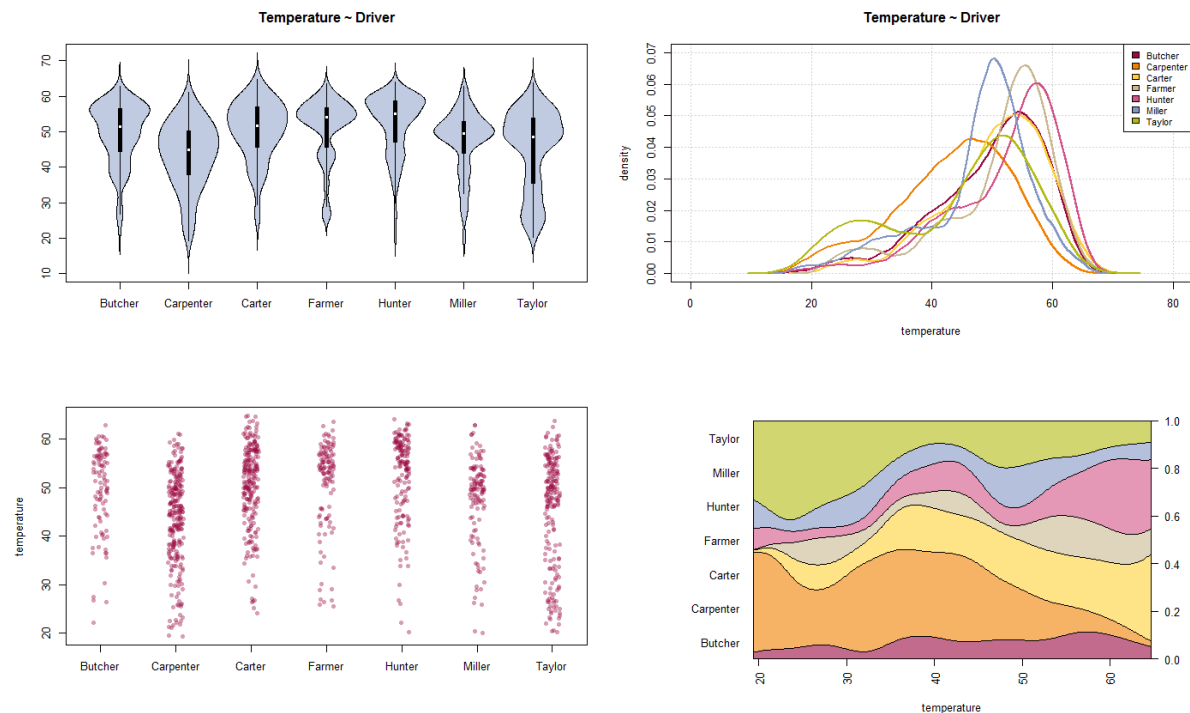
Comparing distributions

How should we compare distributions graphically, moving beyond a simple boxplot? PlotViolin serves the same utility as a side-by-side boxplot, but provides more detail about the single distribution. We started with John Verzani's Violinplot and rewrote it so that it takes exactly the same parameters as the boxplot-function.

Another idea is to plot several densities within the same plot. PlotMultiDens does this while setting the xlim- and ylim-values to an appropriate value, ensuring all density lines are fully visible. For a smaller number of variables, say up to two handfuls, this will be the most direct way to compare their distributions. (Note: For violins this limit lies much higher as they do not overlap and so mutually hide.)

```
PlotViolin(temperature ~ driver, data=d.pizza, col = SetAlpha(hblue,0.5),
           main="Temperature ~ Driver")

PlotMultiDens(temperature ~ driver, data=d.pizza, xlab="temperature",
              main="Temperature ~ Driver", panel.first=grid(),
              col=PalHelsana(), lwd=2 )
```



For small datasets a stripchart might be the best way to plot the data.

The conditional density-plot at the right allows to grasp the proportions within the total density.

```
stripchart(temperature ~ driver, d.pizza, vertical=TRUE,
           method="jitter", pch=16, col=SetAlpha(hred,0.4))

d.frm <- na.omit(d.pizza[,c("temperature", "driver")])
par(las=2, mar=c(4.1, 10.1, 5.1, 5.1))
cdplot(x=d.frm$temperature, y=d.frm$driver, ylab="", xlab="temperature",
       col=SetAlpha(PalHelsana(), 0.6))
```

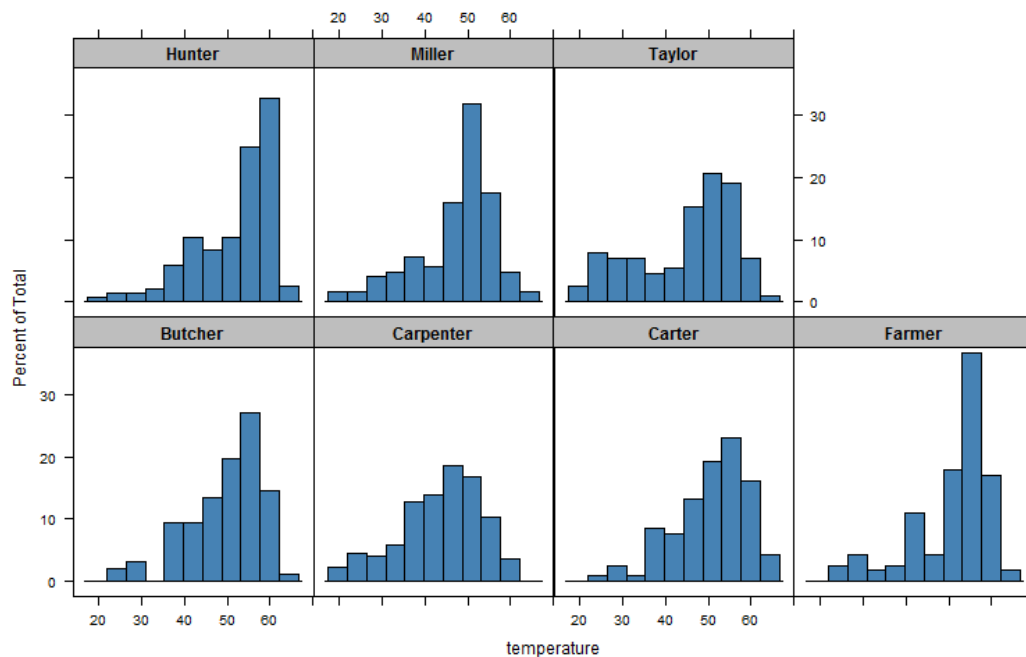
Several distributions with Trellis

The classic way is to spend a full plot for every single variable. There's an interesting link, demonstrating this technique: <http://www.statmethods.net/advgraphs/trellis.html>

```
library(lattice)
trellis.par.set(strip.background = list(col = gray(0.5)), add.text = list(col = 'white'))

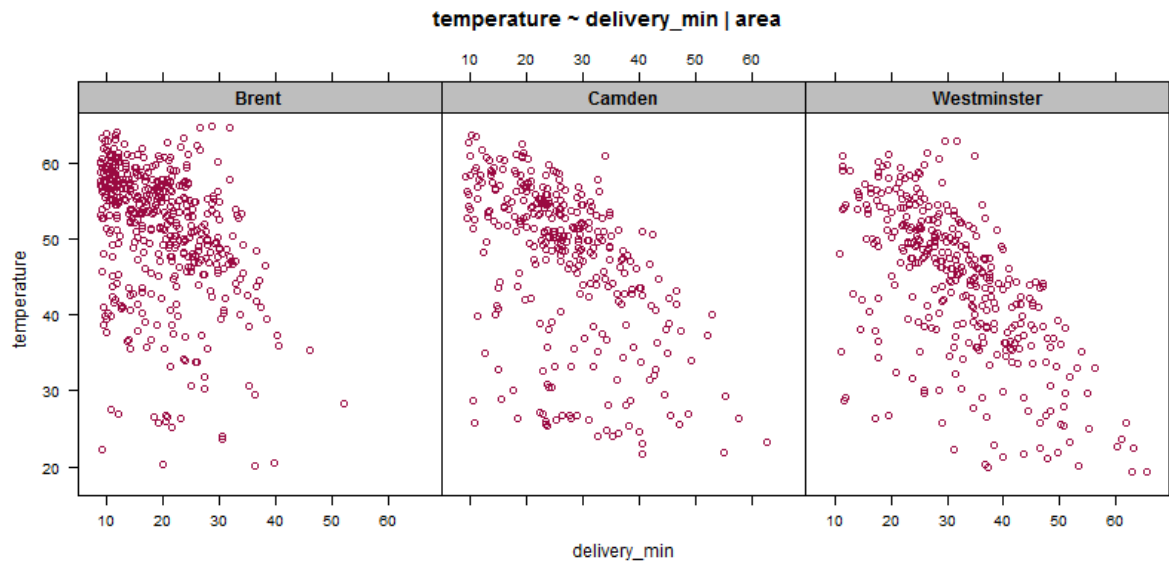
myStripStyle <- function(which.panel, factor.levels, ...) {
  panel.rect(0, -0.5, 1, 1,
    col = "grey",
    border = 1)
  panel.text(x = 0.5, y = 0.25,
    font=2,
    lab = factor.levels[which.panel],
    col = "black")
}

histogram( ~ temperature | driver, data=d.pizza, col="steelblue", strip=myStripStyle)
```



Again here a scatterplot is highly informative.

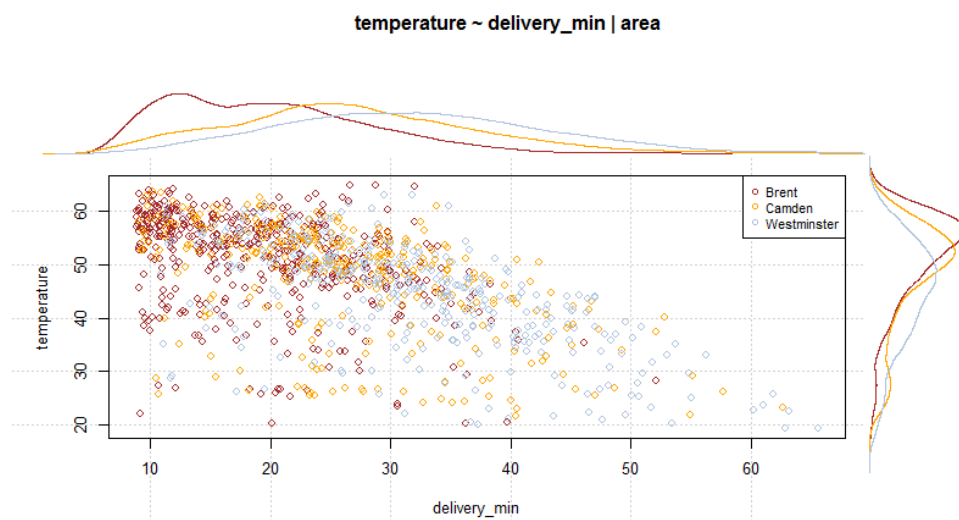
```
xyplot(temperature ~ delivery_min | area, d.pizza, main='temperature ~ delivery_min | area',
  col=hred, strip=myStripStyle)
```



PlotMarDens

This plot shows a scatterplot of two numerical variables temperature and delivery_time, by area. On the margins the density curves of the specific variable are plotted, also stratified by area.

```
PlotMarDens(y=d.pizza$temperature, x=d.pizza$delivery_min, grp=d.pizza$area,
            xlab="delivery_min", ylab="temperature",
            col=c("brown", "orange", "lightsteelblue"), panel.first=grid(),
            main="temperature ~ delivery_min | area" )
```

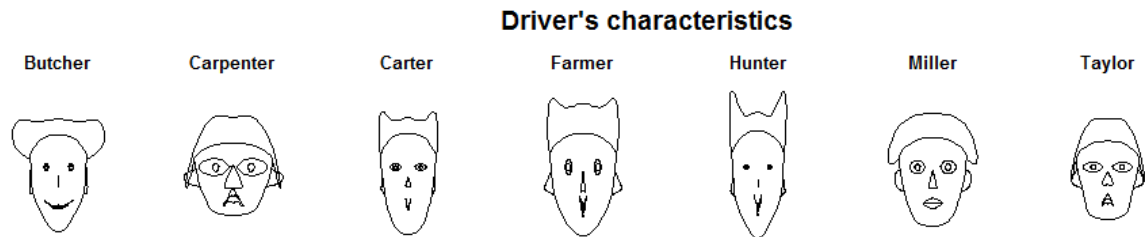


PlotFaces

A nice idea for the concrete representation of your customer's profile is to produce a Chernoff faces plot. The rows of a data matrix represent cases and the columns the variables.

```
m <- data.frame( lapply( d.pizza[,c("temperature","price","delivery_min","wine_ordered","weekday")]
                        , tapply, d.pizza$driver, mean, na.rm=TRUE))

PlotFaces(m, ncol=7, nrow=1, main="Driver's characteristics")
```



Correlations: PlotCorr and PlotWeb

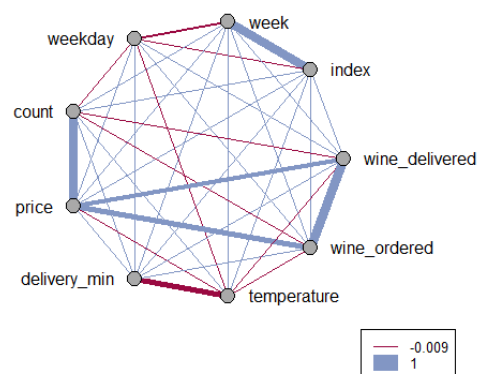
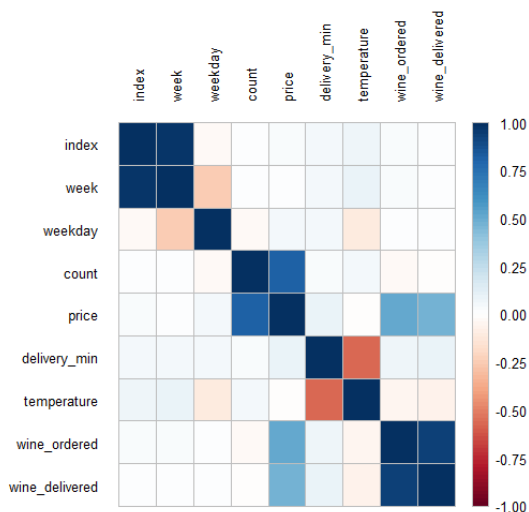
These functions produce a graphical display of a correlation matrix. In the classic matrix representation the cells of the matrix can be shaded or coloured to show the correlation value.

In the right circular representation the correlations are coded in the line width of the connecting lines. Red means a negative correlation, blue a positive one.

```
par(mfrow=c(1,2))
m <- cor(d.pizza[, WhichNumerics(d.pizza)], use="pairwise.complete.obs")

PlotCorr(m, col=PalDescTools("RedWhiteBlue1", 100), border="grey",
        args.colorlegend=list(labels=FormatFix(seq(1,-1,-.25), 2), frame="grey"))

PlotWeb(m, col=c(hred, hblue))
```



Associations with circular plots

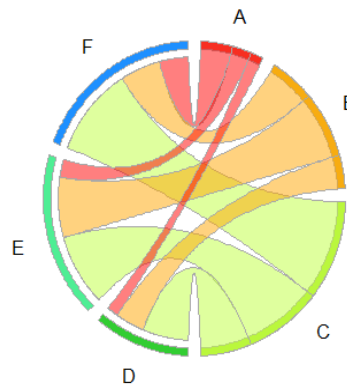
Saw this lately and considered it a promising idea. Still this is, although working, rather experimental code.

```
tab <- matrix(c(2,5,8,3,10,12,5,7,15), nrow=3, byrow=FALSE)
dimnames(tab) <- list(c("A","B","C"), c("D","E","F"))

PlotCirc( tab,
  acol = c("dodgerblue","seagreen2","limegreen","olivedrab2","goldenrod2","tomato2"),
  rcol = SetAlpha(c("red","orange","olivedrab1"), 0.5)
)
```

The table

	D	E	F
A	2	3	5
B	5	10	7
C	8	12	15



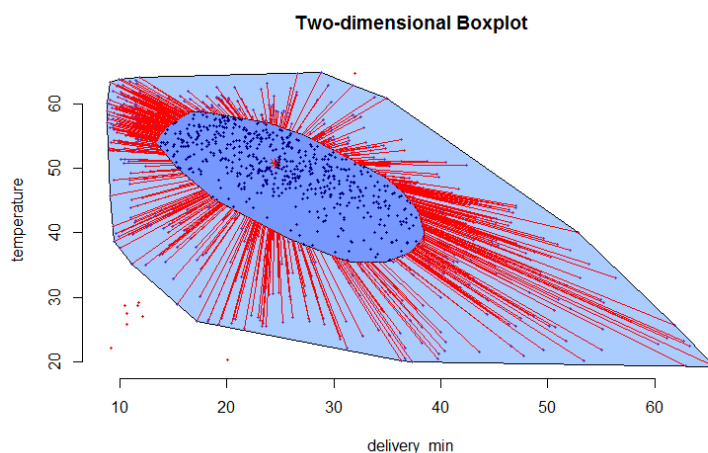
Boxplot on 2 dimensions: PlotBag

This function transposes the boxplot idea in the 2-dimensional space. The points are outliers, the lightblue area is the area within the fences in a normal boxplot and the darkblue area is the inner quartile range. The median is plotted as orange point in the middle.

This code is taken verbatim from Peter Wolf's aplpack package.

```
d.frm <- d.pizza[complete.cases(d.pizza[,c("temperature","delivery_min")]),]

PlotBag(x=d.frm$delivery_min, y=d.frm$temperature, xlab="delivery_min", ylab="temperature",
  main="Two-dimensional Boxplot")
```



PlotPolar (Radarplot)

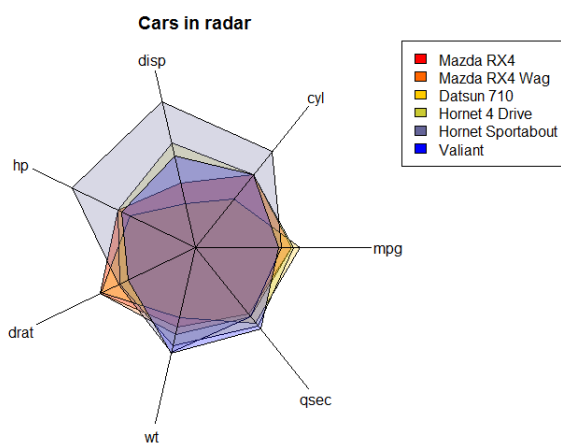
This function produces a polar plot but can also be used to draw radarplots or spiderplots.

A)

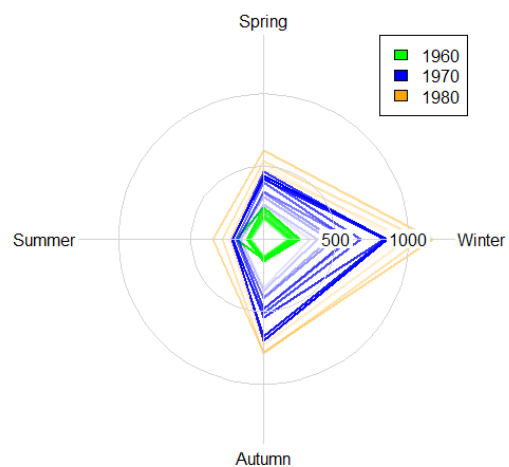
```
d.car <- scale(mtcars[1:6,1:7], center=FALSE)

# let's have a palette with thransparent colors
cols <- SetAlpha(colorRampPalette(c("red","yellow","blue"), space = "rgb")(6), 0.25)

PlotPolar(d.car, type="l", fill=cols, main="Cars in radar")
PolarGrid(nr=NA, ntheta=ncol(d.car), alabels=colnames(d.car), lty="solid", col="black")
legend(x=2, y=2, legend=rownames(d.car), fill=SetAlpha(cols, NA))
```



A)



B)

B)

```
m <- matrix(UKgas, ncol=4, byrow=TRUE)

cols <- c(SetAlpha(rep("green", 10), seq(0,1,0.1)),
          SetAlpha(rep("blue", 10), seq(0,1,0.1)),
          SetAlpha(rep("orange", 10), seq(0,1,0.1)))

PlotPolar(r=m, type="l", col=cols, lwd=2 )
PolarGrid(ntheta=4, alabels=c("Winter","Spring","Summer","Autumn"), lty="solid")

legend(x="topright", legend=c(1960,1970,1980), fill=c("green","blue","orange"))
```

```

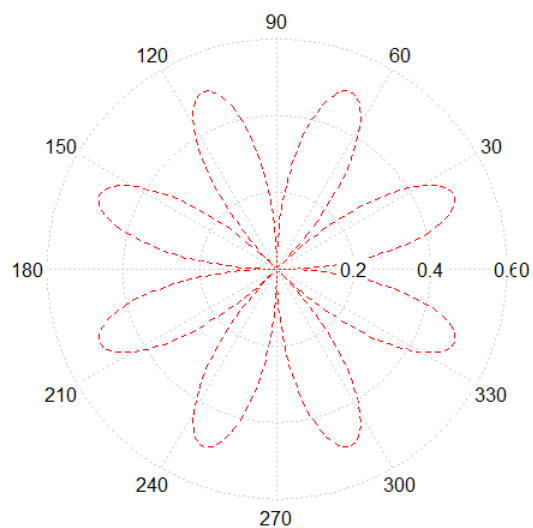
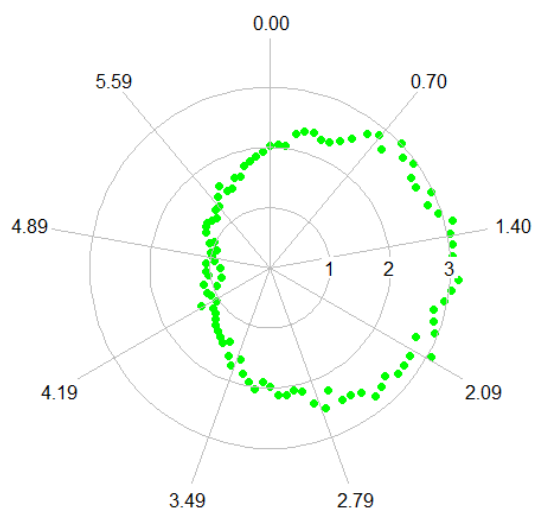
testlen <- c(sin(seq(0, 1.98*pi, length=100)) + 2 + rnorm(100)/10)
# start at 12 o'clock and plot clockwise
PlotPolar(testlen, -(testpos - pi/2), type="p", main="Test Polygon", col="green", pch=16)

PolarGrid(ntheta = rev(seq(0, 2*pi, by=2*pi/9) + pi/2),
          alabels=FormatFix(seq(0, 2*pi, by=2*pi/9),2)[-10], col="grey",
          lty="solid", lblradians=TRUE)

# just because of its beauty
t <- seq(0,2*pi,0.01)

PlotPolar( r=sin(2*t)*cos(2*t), theta=t, type="l", lty="dashed", col="red" )
PolarGrid()

```



Plot Functions

Functions can be plotted a bit more comfortable by means of the function `PlotFct`. The idea is to be able to use the formula interface, for example $x^2 \sim x$, and let the function choose appropriate defaults for the rest. (This would be the best case scenario...;-).

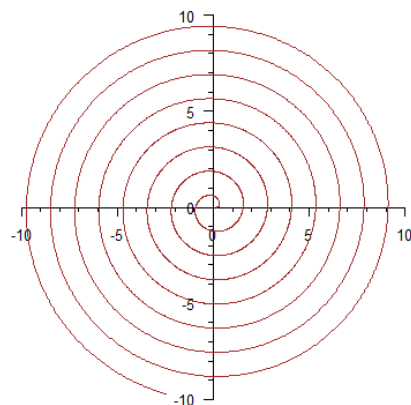
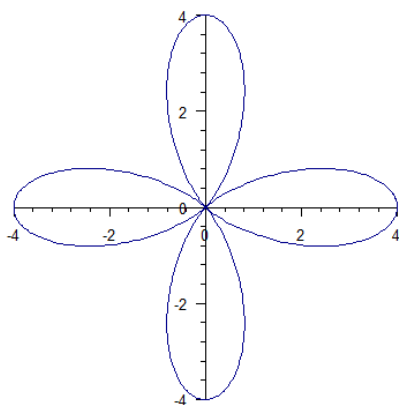
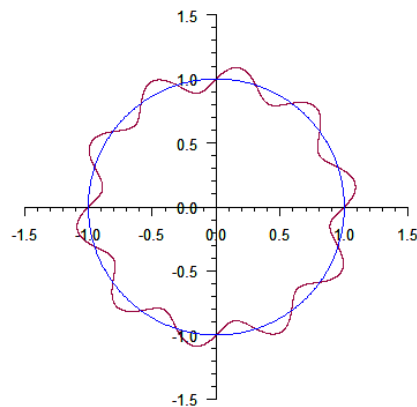
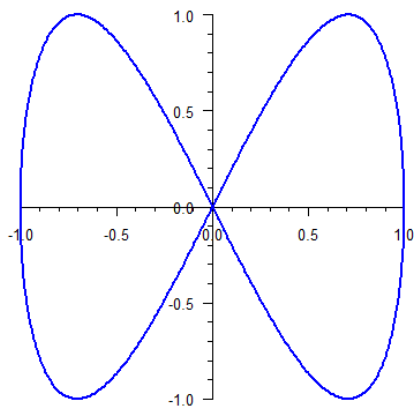
There can as well be further parameters defined for plotting more than one function at once.

```
# get some data
par(mfrow=c(2,2))
PlotFct(sin(2*t) ~ sin(t), from=0, to=2*pi, by=0.01, col="blue", lwd=2)

PlotFct(1+ 1/10 * sin(10*x) ~ x, polar=TRUE, from=0, to=2*pi, by=0.001, col=hred)
PlotFct(sin(x) ~ cos(x), polar=FALSE, from=0, to=2*pi, by=0.01, add=TRUE, col="blue")

# lemniscate of Bernoulli
PlotFct((2*a^2*cos(2*t))^2 ~ t, args=list(a=1), polar=TRUE, from=0, to=2*pi+0.1, by=0.01,
        col="darkblue")

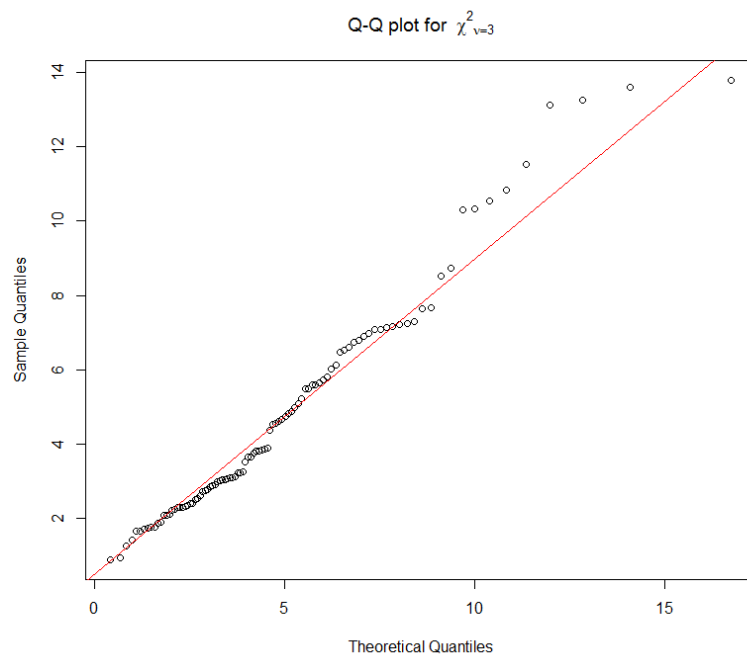
# evolving circle
PlotFct(a*(sin(t) - t*cos(t)) ~ a*(cos(t) + t*sin(t)), args=list(a=0.2), from=0, to=50, by=0.01,
        col="brown")
```



PlotQQ

This is a short wrapper for plotting QQ-plots with other than normal distributions. A qqline is inserted.

```
# get some data
PlotQQ(z, function(p) qchisq(p, df=5), args.qqline=list(col=2, probs=c(0.1,0.6)),
      main=expression("Q-Q plot for" ~{chi^2}[nu == 3]))
```



PlotTreemap

This function produces a treemap.

```
# get some data
data(GNI2010, package="treemap")
gn <- GNI2010[,c("iso3", "population", "continent", "GNI")]
gn <- gn[gn$GNI!=0,]

# define a color
gn$col1 <- SetAlpha("steelblue", LinScale(gn$GNI, newlow=0.1, newhigh=0.6))

b <- PlotTreemap(x=gn$population, grp=gn$continent, col=gn$col1, labels=gn$iso3,
  main="Gross national income (per capita) in $ per country in 2010",
  labels.grp=NA, cex=0.7)

# get the midpoints
mid <- do.call(rbind, lapply(lapply(b, "[", 1), data.frame))

# and write the continents' text
DrawBoxedText(x=mid$grp.x, y=mid$grp.y, labels=rownames(mid), cex=1.5, bold=TRUE,
  border=NA, col=SetAlpha("white",0.7) )
```

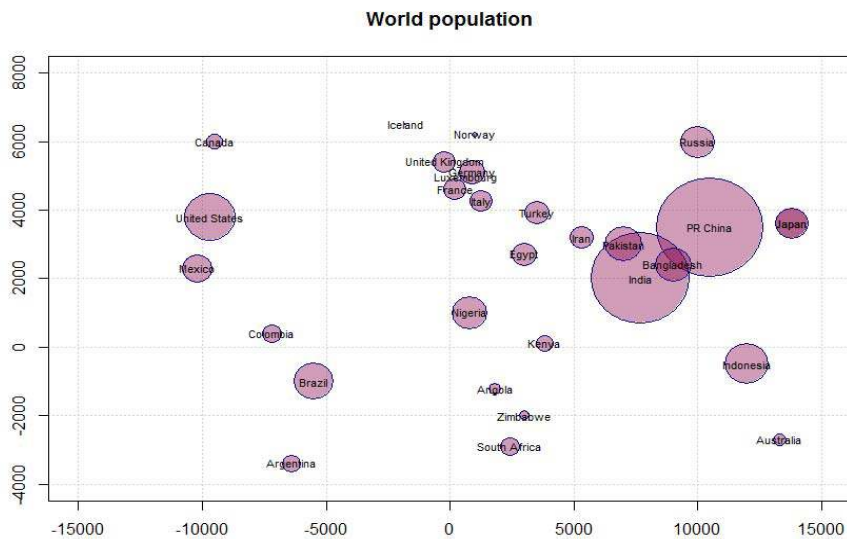
Gross national income (per capita) in \$ per country in 2010



PlotBubble

Bubbles can actually easily be produced with the standard plot function. This function here helps you defining appropriate axis limits.

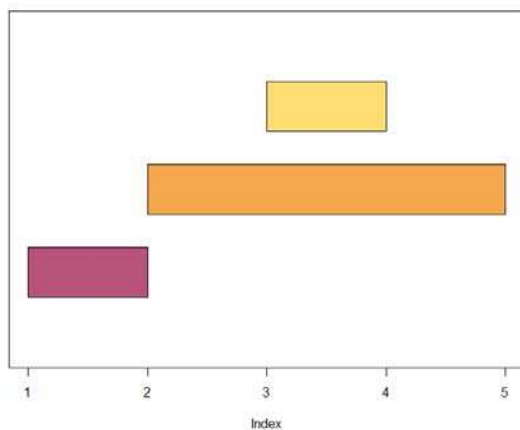
```
PlotBubble(d.world$x, d.world$y, area=d.world$pop/90, col=SetAlpha("deeppink4",0.4), border="darkblue",
           xlab="", ylab="", panel.first=grid(), main="World population")
text(d.world$x, d.world$y, labels=d.world$country, cex=0.7, adj=0.5)
```



PlotHorizBar

This is a simple function for plotting flowing horizontal or vertical bars.

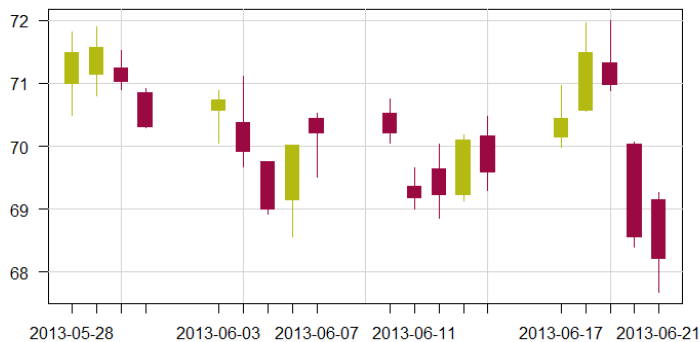
```
PlotHorizBar(from=c(1,2,3), to=c(2,5,4), grp=c(1,2,3), col=PalHelsana()[1:3])
```



PlotCandlestick

This plot is used primarily to describe price movements of a security, derivative or currency over time. Candlestick charts are a visual aid for decision making in stock, foreign exchange, commodity, and option trading.

```
example(PlotCandlestick)
PlotCandlestick(x=as.Date(rownames(nov)), y=nov, border=NA, las=1, ylab="")
```

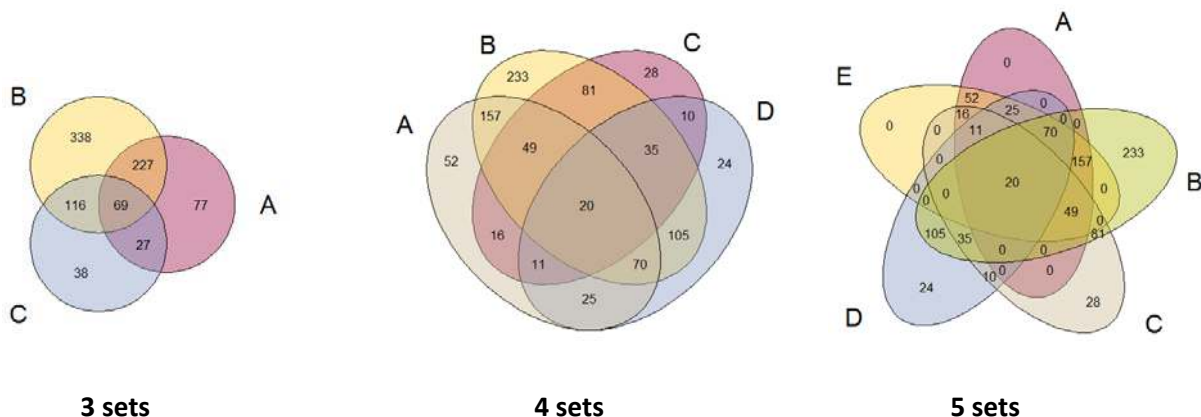


Venn plots

In rare cases one might want to plot a Venn diagram. This function does this for up to 5 datasets using the simple proposed geometric representations.

(For more than 5 datasets the Venn representation loses its simplicity and other plot types become more adequate.)

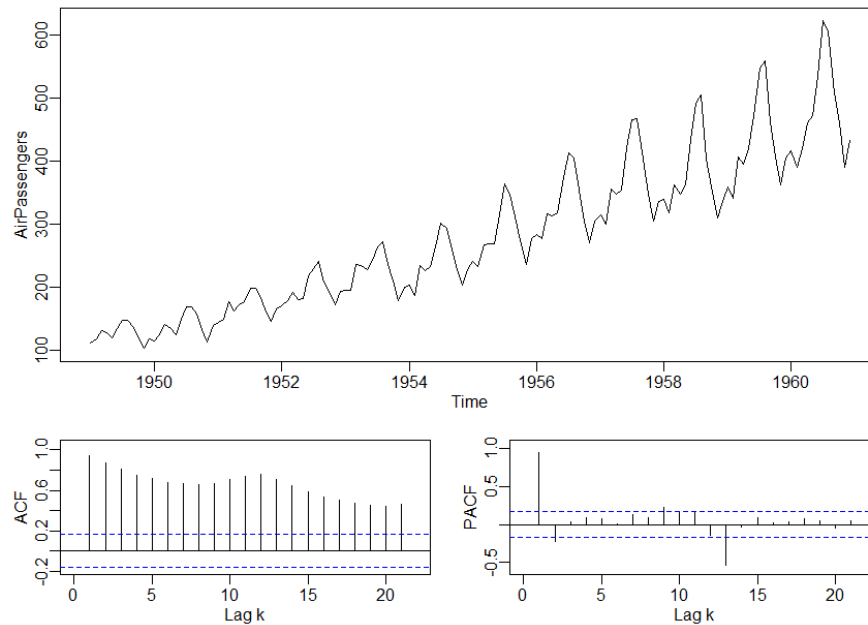
```
example(PlotVenn)
PlotVenn(x=x[1:3], col=SetAlpha(c(PalHelsana()[c(1,3,6)]), 0.4))
PlotVenn(x=x[1:4], col=SetAlpha(c(PalHelsana()[c(1,3,6,4)]), 0.4))
PlotVenn(x=x[1:5], col=SetAlpha(c(PalHelsana()[c(1,3,6,4,7)]), 0.4))
```



PlotACF

This produces a combined plot of a time series and its autocorrelation and partial autocorrelation

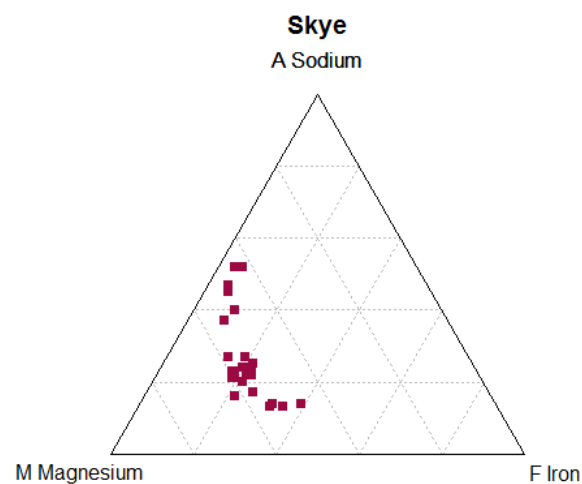
```
PlotACF(AirPassengers)
```



PlotTernary

This produces a ternary or triangular plot.

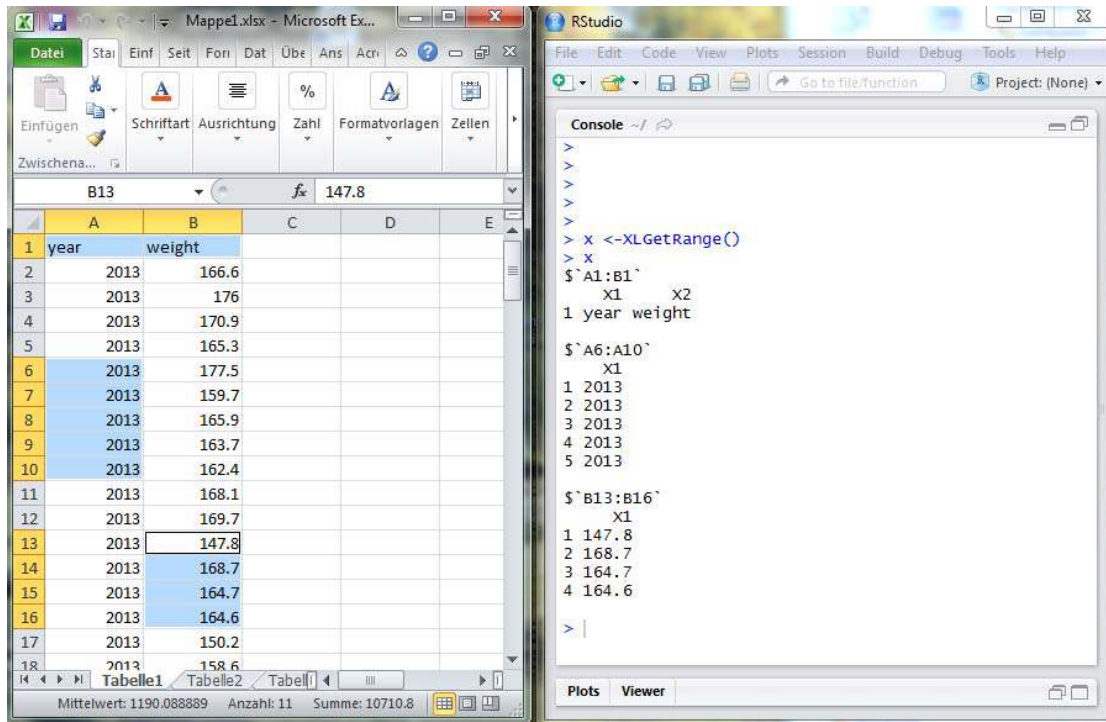
```
data(Skye, package="MASS")
PlotTernary(Skye[c(1,3,2)], pch=15, col=hred, main="Skye",
            lbl=c("A Sodium", "F Iron", "M Magnesium"))
```



Import data via Excel

The function `XLGetRange` allows a quick import of data from an Excel-Sheet. The user can either specify a number of cell-references (including a path- and filename) or just select the regions which are to be imported.

The following command will return a list with the contents of the selected cell ranges.



`XLView(d.frm)` can be used to view a data.frame `d.frm` in Excel.

Import SAS datalines

The function `ParseSASDatalines` can be used to import the SAS data like the following:

```
sas <- "
data FatComp;
input Exposure Response Count;
label Response='Heart Disease';
datalines;
  0 0 6
  0 1 2
  1 0 4
  1 1 11
;"
```

```
(FatComp <- ParseSASDatalines(sas))
```

```
Exposure Response Count
1      0      0      6
2      0      1      2
3      1      0      4
4      1      1     11
```

String functions

As the string functions are highly scattered, they sometimes can hardly be found right from the start. This overview might be helpful:

<code>read.lines</code>	Read a text file
<code>cat</code>	Print strings
<code>paste, %c%</code>	Concatenate strings
<code>nchar</code>	Number of characters in a string
<code>substr</code>	Extract or replace substrings in a character vector
<code>StrLeft, StrRight</code>	Same as <code>substr</code> , but with option to get the substring from the right end
<code>match, pmatch</code>	Searching parts of one string in another
<code>charmatch</code>	Partial string matching <code>charmatch</code> seeks matches for the elements of its first argument among those of its second.
<code>grepl</code>	Check if a string contains another string.
<code>grep, regexpr</code>	Searching values in a character vector by means of regular expressions.
<code>sub, gsub</code>	Substitute a part of the string with another
<code>StrVal</code>	Extract numeric values out of a string
<code>StrPos</code>	Finds the first position of one string in another string
<code>StrTrim</code>	Delete white spaces from a string
<code>strtrim</code>	Trim character strings to specified display widths.
<code>StrTrunc</code>	Truncate string on a given length and add ellipses if it really was truncated
<code>StrChop</code>	Split a string by a fixed number of characters.
<code>strsplit</code>	Split the elements of a character vector <code>x</code> into substrings according to the matches to substring split within them.
<code>StrDist, adist</code>	Compute distances between strings
<code>AscToChar,</code> <code>CharToAsc</code>	Converts ASCII codes to characters and vice versa
<code>abbr, StrAbbr</code>	Abbreviates a string
<code>StrCap</code>	Capitalize the first letter of a string
<code>tolower, toupper</code>	convert upper-case characters in a character vector to lower-case, or vice versa
<code>StrRev</code>	Reverse a string
<code>StrCountW</code>	Count the words in a string
<code>StrIsNumeric</code>	Checks whether a string contains only numeric data
<code>chartr</code>	Translate characters in character vectors, in particular from upper to lower case or vice versa.
<code>sprintf</code>	Format a numeric value as a string

References

Agresti, A. (2002) Categorical Data Analysis. John Wiley & Sons.

Dalgaard, P. (2008) Introductory Statistics with R (2. Aufl.), London, UK: Springer.

Wollschläger, D. (2010, 2012) Grundlagen der Datenanalyse mit R, Springer, Berlin.

Tufte, Edward R (2001) [1983], The Visual Display of Quantitative Information (2nd ed.), Cheshire, CT: Graphics Press